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# ABSCISIC ACID IN *AEGILOPS KOTSCHYI* CARYOPSES

JUDITH WURZBURGER and YA'ACOV LESHEM

Department of Life Sciences, Bar-Ilan University, Ramat-Gan, Israel

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**Key Word Index**—*Aegilops kotschy*; Gramineae; caryopsis; abscisic acid; germination inhibitor.

The glumes and hulls of *Aegilops kotschy* (Gramineae) contain material which inhibits the germination of the seeds of this plant [1] and lettuce [1–2]. An extract also accelerates leaf abscission in cotton seedlings [2] and inhibits GA<sub>3</sub>-promoted reducing sugar production [2–3]. An attempt was made to determine the amount of the inhibitor present in the caryopses and identify it as ABA.

It was found that an inhibitor exists in the acidic ethyl acetate fraction which was prepared from the two types of caryopses. This inhibitor repressed the production of GA<sub>3</sub>-promoted reducing sugar. The  $R_f$  values of the acidic ethyl acetate fraction of each caryopsis type corresponding to the  $R_f$  of the synthetic marker ABA were eluted from the chromatograms. After methylation, the elutes were subjected to GLC. It was found that the extract of the caryopses had a peak at the  $R_i$  of 3.0 min which exactly corresponded to  $R_i$  of the marker (the *cis-trans* isomer). It thus appears that *cis-trans*-ABA is present in both types of caryopses, its concentration being 2.5 times higher in the smaller (6.2 ng/g dry wt) than in the larger one (2.5 ng/g dry wt), this probably being a contributing factor to the more marked dormancy of the former [3–4].

## EXPERIMENTAL

**Plant material.** *Aegilops kotschy* spikelets were collected in the Northern Negev of Israel in the summer of 1971. After

dehulling, large and small caryopses were investigated separately.

**Extraction.** 5 g dehulled smaller and larger caryopses were homogenized in cold 80% MeOH. After shaking the extract for 24 hr in the cold, it was filtered and centrifuged for 20 min at 10000 g. The MeOH in the supernatant was removed under vac at 35° and the pH of the aq. layer adjusted to 8.5 with 5% NaHCO<sub>3</sub>. This was extracted 4× light petrol (30°–40°) and then 4× EtOAc. The pH was then brought to 3.0 with 1 N HCl, and again extracted 4× EtOAc. The latter fractions were combined and evaporated to dryness.

**Chromatography.** Samples of the EtOAc fraction (equivalent to 1 g dry wt) were separated on Whatman no. 1 paper and with PrOH–NH<sub>4</sub>OH–H<sub>2</sub>O (10:1:1). The barley endosperm test for gibberellins and ABA [5] revealed the presence of an inhibitory zone at  $R_f$  corresponding to that of the synthetic marker (Sigma). The zones were eluted with MeOH and methylated with CH<sub>3</sub>N<sub>2</sub> and separated by GLC column 1.8 m × 0.3 cm with 1.5% QF<sub>1</sub> on Gas Chrom Q, 60–80 mesh with N<sub>2</sub> (26 ml/min) at 200°.

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# A NEW NAPHTHAQUINONE FROM *TABEBUIA GUAYACAN*

GARY D. MANNERS and LEONARD JURD

Western Regional Research Laboratory, Agricultural Research Service, U.S. Department of Agriculture, Berkeley, California 94710, U.S.A.

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**Key Word Index**—*Tabebuia guayacan*; Bignoniaceae; 2,3-di(3,3-dimethylallyl)-1,4-naphthaquinone;  $\alpha$ - and  $\beta$ -lapachones.

**Plant:** *Tabebuia guayacan* Hemsley. **Source:** Panama. **Previous work:** An early report [1] of lapachol in the wood of *T. guayacan* is the only reported investigation prior to the current work [2] in this laboratory. Other *Tabebuia* species have yielded several naphthaquinones, anthraquinones and prenylnaphthalene dimers [3–5]. **Present work.** Hammermilled heartwood was extracted successively with petrol bp 30–60°, Et<sub>2</sub>O Me<sub>2</sub>CO and MeOH. Recrystallization of cold petrol insolubles

yielded lapachol (mp 136–137°) and preparative column chromatography of the filtrate (deactivated Si gel, C<sub>6</sub>H<sub>6</sub>) yielded a new compound as yellow needles (MeOH) mp 72–73°. Found: C, 81.2; H, 7.51 [Calc. for C<sub>20</sub>H<sub>22</sub>O<sub>2</sub>: C, 81.6; H, 7.53]. UV  $\lambda_{\max}^{1:1\text{OH}}$  (log  $\epsilon$ ): 327 (3.25), 268 (4.20), 260 (4.19), 245 (4.25), ~243 (4.26) nm. IR:  $\nu_{\max}^{\text{Nujol}}$  1660, 1615, 1590, 1460, 1290, 1280, 1160, 1100, 950, 345, 720 cm<sup>-1</sup>. The 100 MHz NMR in CDCl<sub>3</sub> showed two vinyl gem dimethyl groups (s at  $\delta$ 1.70 and  $\delta$ 1.80), 2 equivalent